



## Biosynthesis of silver nanoparticles by using onion (*Allium cepa*) extract and study antibacterial activity

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### Abstract

In the present research program, cost effective and environment friendly silver nanoparticles were synthesized using the onion (*Allium cepa*) extract as the reducing and capping agent. The nanoparticles were characterized using UV-visible, FT-IR, XRD, AFM and SEM methods. The surface plasmon resonance peaks in absorption spectra for silver colloidal solution showed that the absorption maximum range was at 380-440nm. The functional biomolecules such as carboxyl groups present in the seaweed responsible for the silver nanoparticles formation were characterized by FT-IR. The XRD results suggested that the crystallization of the bio-organic phase occurs on the surface of the silver nanoparticles or vice versa. The broadening of peaks in the XRD patterns was attributed to particle size effects and the average particles size about 25.17nm. The silver nanoparticles examined by AFM have an average diameter about 28.4nm and an average height 21.9nm. We recognize a very good agreement between the two results: a XRD-size corresponds effectively to a size of AFM. The silver nanoparticles synthesized by the help of onion extract were scanned using SEM. It reveals that a silver nanoparticle seems to be spherical in morphology. The silver nanoparticles synthesis by onion extracts was showed complete antibacterial activity against *Escherichia coli*, *Proteus mirabilis*, *Klebsiella oxytoca*, *Streptococcus* sp., *Enterobacter cloacae*, *Bacillus* sp., *Pseudomonas aeruginosa* and *Staphylococcus aureus* were collected from diagnosis laboratory, Al-Numan hospital, Baghdad. The study revealed that the silver nanoparticles synthesis by onion extracts could be as a therapeutic agent for human microbial infections.

Keywords: Silver nanoparticles, Pathogenic bacteria, Onion, Antimicrobial activity.

### Introduction

Nanotechnology is a rapidly growing science of producing and utilizing nano-sized particles. A number of approaches are available for the synthesis of silver nanoparticles, such as thermal decomposition (Navaladian *et al.*, 2007), electrochemical (Starowicz *et al.*, 2006) microwave assisted process (Sreeram *et al.*, 2008) and green chemistry (Begum *et al.*, 2009). Many of the nanoparticle synthesis or production methods of nanoparticles involve the use of hazardous chemicals, low material conversions and high energy requirements. So, a growing need to develop an environmentally friendly process for nanoparticle synthesis without using toxic chemicals is gaining importance. Biosynthetic methods employing either microorganisms or plant extracts have emerged as a simple and viable alternative to chemical synthetic procedures and physical methods. Several microorganisms, such as bacteria, fungi and yeasts, have come up as nano

factories, synthesizing metal nanoparticles of Ag and Au. However, the use of plants for the fabrication of nanoparticles has drawn attention, because of its rapid, economical, eco-friendly protocol, and it provides a single step technique for the biosynthesis process (Huang *et al.*, 2007). Biological approaches using microorganisms and plants or plant extracts for metal nanoparticle synthesis have been suggested as valuable alternatives to chemical methods. An important branch of biosynthesis of nanoparticles is the application of plant extract to the biosynthesis reaction. Synthesis of quasi spherical silver nanoparticles used a purified apiin compound, extracted from henna leaf at ambient conditions (Kasthuri *et al.*, 2009). Using green tea, *C. sinensis* extracts, as a reducing and stabilizing agent, gold nanoparticles and silver nanostructures were produced in aqueous solution at ambient conditions (Nestor *et al.*, 2008). Plant extracts from live alfalfa, the broths of lemongrass, geranium leaves and

others have served as green reactants in Ag NP synthesis. The reaction of aqueous AgNO<sub>3</sub> with an aqueous extract of leaves of a common ornamental geranium plant, *Pelargonium graveolens*, gave Ag NPs after 24hr (Shankar *et al.*, 2003). Silver nanoparticles ranging from 55 to 80nm in size, and triangular or spherical gold nanoparticles, were fabricated using the novel sundried biomass of *Cinnamomum canphora* leaf (Huang *et al.*, 2007). A simple procedure applying *Aloe vera* leaf extract has been used for the synthesis of gold nanotriangle and spherical silver nanoparticles. *Aloe vera* extract showed more spherical silver nanoparticles with increasing the amount of added extract (Chandran, *et al.*, 2006). Silver nanoparticles were successfully synthesized using the latex of *Jatropha curcas*. The plant, *Jatropha curcas*, is commercially important, as bio-diesel is extracted from its seeds on an industrial scale. Crude latex was obtained by cutting the green stems of *J. curcas* plants (Bar *et al.*, 2009). In another research work, biosynthesis of silver nanoparticles was also conducted using Cycas leaf extract (Jha and Prasad, 2010). Cycas is rich in flavonoids, broadly belonging to the class of phenolic compounds.

With the emergence and increase of microbial organisms resistant to multiple antibiotics, and the continuing emphasis on health-care costs, many researchers have tried to develop new, effective antimicrobial reagents free of resistance and cost. Such problems and needs have led to the resurgence in the use of Ag-based antiseptics that may be linked to broad-spectrum activity and far lower propensity to induce microbial resistance than antibiotics (Jones *et al.*, 2004). The antibacterial effects of Ag salts have been noticed since antiquity (Silver and Phung, 1996) and Ag is currently used to control bacterial growth in a variety of applications, including dental work, catheters, and burn wounds (Catauro *et al.*, 2004; Crabtree *et al.*, 2003).

The present work has focused on the development of the easy synthesis of silver nanoparticles by an environmentally friendly procedure. Onion (*Allium cepa*) extract was used for the silver nanoparticles synthesis, and evaluation of their antibacterial activity against various human pathogenic bacteria.

### Materials and Methods

**Collection of pathogens:** We are collected multiple antibiotic-resistant isolates, which included *Escherichia coli*, *Proteus mirabilis*, *Klebsiella oxytoca*, *Streptococcus sp.*, *Enterobacter cloacae*, *Bacillus sp.*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* used for the antimicrobial

activity from microbiology diagnosis laboratory, Al-Numan hospital.

**Synthesis of silver nanoparticles:** For the synthesis of silver nanoparticles, we used onion (*Allium cepa*) as an extract for reducing and capping agent which is purchased from market and silver nitrate (AgNO<sub>3</sub>) from Merck limited, India. Onion extract was prepared by taking 25g of thoroughly washed and finely crushed onion mixed with 100ml deionized water in 500 ml of Erlenmeyer flask and then boiling the mixture for 10 min before finally decanting it. For the reduction of Ag<sup>+</sup> ions, 5ml of onion extract was mixed to 50 ml of 0.1mM aqueous of AgNO<sub>3</sub> solution drop wise with constant stirring at 50-60°C and observe the color change.

**Characterization of silver nanoparticles:**

**UV-Vis Spectra analysis:** The reduction of pure Ag<sup>+</sup> ions was monitored by measuring the UV-Vis spectrum of the reaction medium after 30 min. UV-Vis spectrophotometer is procured from Shimadzu. A small aliquot of the sample was taken for UV-Vis spectrum analysis (350-750 nm). The maximum absorbance spectrum of As-Ag nanoparticles was observed at 455 nm.

**Fourier Transform Infra-Red Spectroscopy (FT-IR):** FT-IR measurements were carried out using Perkin (8300 FT-IR Shimadzu Spectrophotometer) the range from 4000 cm<sup>-1</sup> to 400 cm<sup>-1</sup>. After complete reduction of AgNO<sub>3</sub> ions by onion (*Allium cepa*) extract, the mixture was centrifuged at 10000 rpm for 10 min to remove protein or other bioorganic compounds that were present in the solution. The silver nanoparticles pellet obtained was air dried. The dried nanoparticles were mixed with the potassium bromide (KBr) to made thin pellets and were used for FT-IR analysis in transmittance mode.

**X-Ray Diffraction (XRD) analysis:** Resulting solution of the developed nanoparticles of silver was centrifuged at 10,000 rpm for 30 min. The solid residues of Ag NPs were washed twice with double distilled water and then dried at 80°C to obtain powder Ag NPs used for X-ray powder diffraction measurements. The powder X-ray diffraction (XRD) patterns were recorded on (Shimadzu XRD-6000) with copper radiation (Cu K<sub>α</sub>, 1.5406 Å) at 40kV and 30mA.

**SEM Analysis of Silver Nanoparticles:** Scanning Electron Microscopic (SEM) analysis was done using (Inspect S 50) SEM machine. Thin films of the sample were prepared on a carbon coated copper grid by just dropping a very small amount of the sample on the grid.

**AFM Analysis of Silver Nanoparticles:** Atomic Force Microscopy (AFM) (SPHWSY) was used to obtain the surface topography and the average particle size.

**Determination of antimicrobial activity:**

Antibacterial activity of the silver nanoparticles by using onion (*Allium cepa*) extract was evaluated by the cup plate agar diffusion method (Srinivasan *et al.*, 2001). The bacterial cultures were adjusted to 0.5 McFarland turbidity standards and inoculated onto Mueller Hinton agar (MHA, Oxoid) plates. A sterile cork borer was used to make 2 well (6mm in diameter) on the MHA plates. Aliquots of 100 $\mu$ l of silver nanoparticles solution were applied in one of the wells in the culture plates previously seeded with the test organisms. The cultures were incubated at 37°C for 24hrs. A well was made in each of the culture plates and filled with 100 $\mu$ l of onion extract as a control. Antimicrobial activity was determined by measuring the zone of inhibition around each well (excluding the diameter of the well). For each extract, three replicate trials were conducted against each organism.

### Results and Discussion

The reduction of Ag<sup>+</sup> into Ag-NPs during exposure to water extract of *Allium cepa* leaf was able to be followed by the color change. The fresh suspension of *A. cepa* was yellow. However, after the addition of AgNO<sub>3</sub> and stirring for one hour at room temperature, the emulsion turned brown. The color changes in aqueous solutions are due to the surface-plasmon resonance (SPR) phenomenon (Figure 1A and B). The result obtained in this investigation is interesting because it can serve as a foundation in terms of identification of potential forest plants for synthesizing Ag-NPs.



Figure (1): Photograph of *Allium cepa* (A) and silver/ *A. cepa* (B) emulsions after one hour of stirring time.

UV-Vis Spectrophotometry: The Formation of metal nanoparticles by reduction of the aqueous metal ions during exposure of onion (*Allium cepa*) extract may be easily followed by UV-Vis spectroscopy (UV-Shimadzu spectrophotometer). UV-Vis absorption spectrum of silver nanoparticles in the presence of onion extract is shown in Figure (2). The surface plasmon resonance peaks in absorption spectra for silver colloidal solution showed that the absorption maximum range was at 380- 440nm, suggesting that the nanoparticles were dispersed in the aqueous solution with no evidence for aggregation in UV-Vis absorption spectrum Figure (2).

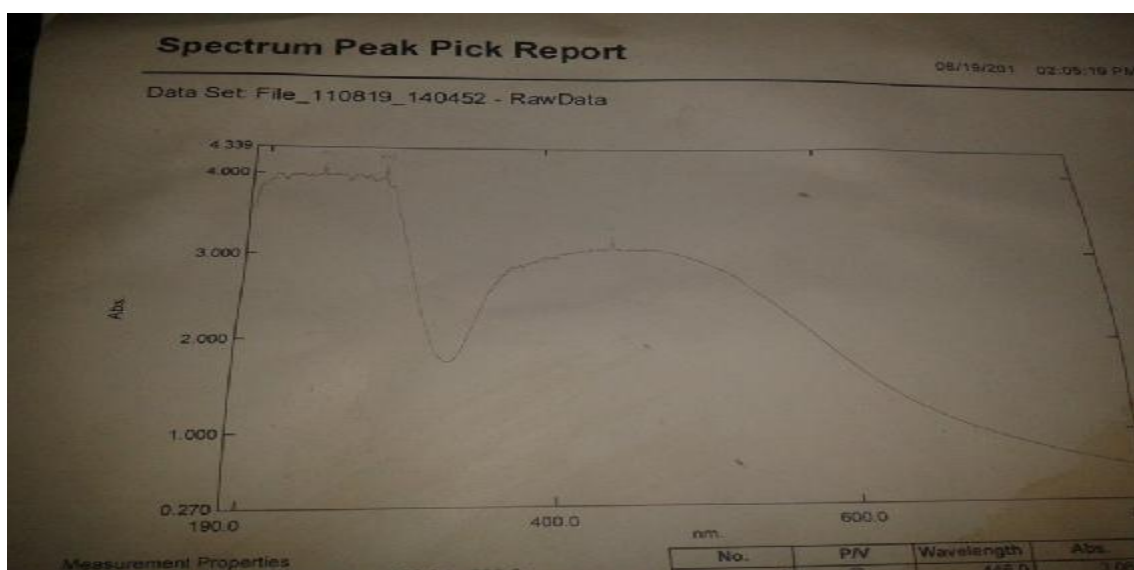


Figure (2): UV-Vis absorption spectra of Silver nanoparticles synthesized by exposure of onion broth with 0.1mM silver nitrate.

**Fourier Transform Infra-Red Spectroscopy:** The results of FTIR analysis of this study show different stretches of bonds shown at different peaks; 3327.21—N—H stretch, 1641.42 —C=C, and 1211.30—C=O. Figure (3) shows the peaks near 3440 $\text{cm}^{-1}$ , and 2968 $\text{cm}^{-1}$  assigned to OH stretching and aldehydic C—H stretching, respectively. The weaker band at 1629 $\text{cm}^{-1}$  corresponds to amide I arising due to carbonyl stretch in proteins. The peak at 1051 $\text{cm}^{-1}$  corresponds to C—N stretching vibration of the amine. The peak near 1743 $\text{cm}^{-1}$  corresponds to C=C stretching (non-conjugated). The peak near 866  $\text{cm}^{-1}$  assigned to C=CH<sub>2</sub> and the peaks near 678 $\text{cm}^{-1}$  and 638 $\text{cm}^{-1}$  assigned to CH out of plane bending vibrations are substituted ethylene systems —CH=CH. FTIR spectra of silver nanoparticles exhibited prominent peaks at 1641 and 1382 $\text{cm}^{-1}$ . The spectra showed sharp and strong absorption band at 1641 $\text{cm}^{-1}$  assigned to the stretching vibration of (NH) C=O group (Barbara *et al.*, 2004).

**Powder X-ray diffraction:** Figure (4) shows the XRD patterns of Ag-NPs. The X-ray diffraction patterns of the Ag-NPs synthesized by using AgNO<sub>3</sub> and using the onion (*Allium cepa*) extract as the reducing and capping agent are shown in Fig. 4. All the reflections correspond to pure silver metal with face centered cubic symmetry. The reflections were indexed as (111), (200) and (220) with the corresponding 2 $\theta$  values of 38.128, 44.315 and 64.468 respectively (JCPDS 04-0783). The intensity of peaks reflected the high degree of crystallinity of the silver

nanoparticles. However, the diffraction peaks were broad indicating that the crystallite size is very small. The average particle size of Ag-NPs can be calculated using the Debye - Scherrer equation (Langford and Wilson, 1978).

$D = K \lambda / \beta \cos \theta$ , where K is the Scherrer constant with value from 0.9 to 1 (shape factor),  $\lambda$  is the X-ray wavelength, 1.5418 Å,  $\beta$  is the width of the XRD peak at half-height and  $\theta$  is the Bragg angle and D is the grain size. From the Scherrer equation, the average crystallite size of Ag-NPs is (25.17nm).

**SEM analysis of Silver nanoparticles:** The silver nanoparticles synthesized by the help of onion extract were scanned by SEM as shown in Figure (5). It reveals that silver nanoparticles seem to be spherical in morphology and particles form cluster. It is easy to notice that the examined particles consist of a number of smaller objects of a few micrometers in size. However, we did not manage to examine the structure of the observed nanoparticles because of difficulties connected with getting higher magnification. In Figure (6), a standard EDX spectrum recorded on the examined sample is shown. In the middle part of the presented spectrum a strong peak located at 3 KV. This maxima is directly related to the silver characteristic line L. The maximum located on the left part of the spectrum at 0.2 kV clearly comes from carbon. Quantitative analysis proved high silver contents (100%) in the examined samples the result shown in Table (1).

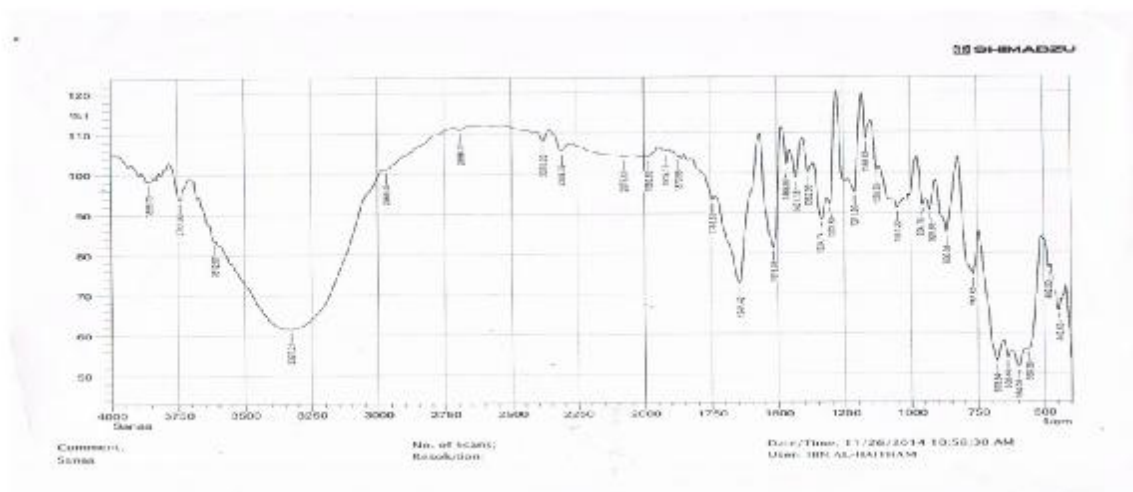


Figure (3): Fourier transform Infra-Red spectroscopy image of silver nanoparticles synthesized by exposure of onion broth with 0.1mM silver nitrate.



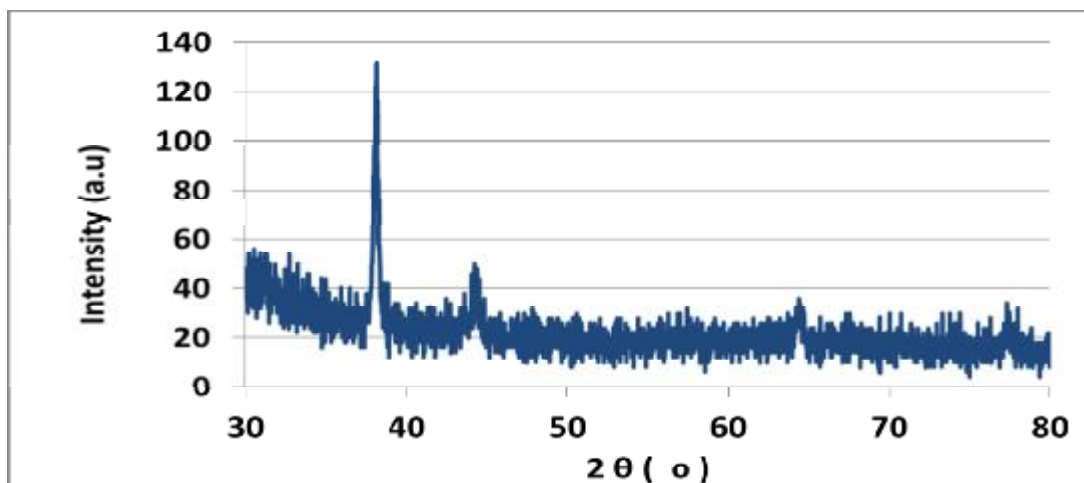


Figure (4): X ray diffraction of Silver nanoparticles synthesized by exposure of onion broth with 0.1mM silver nitrate.

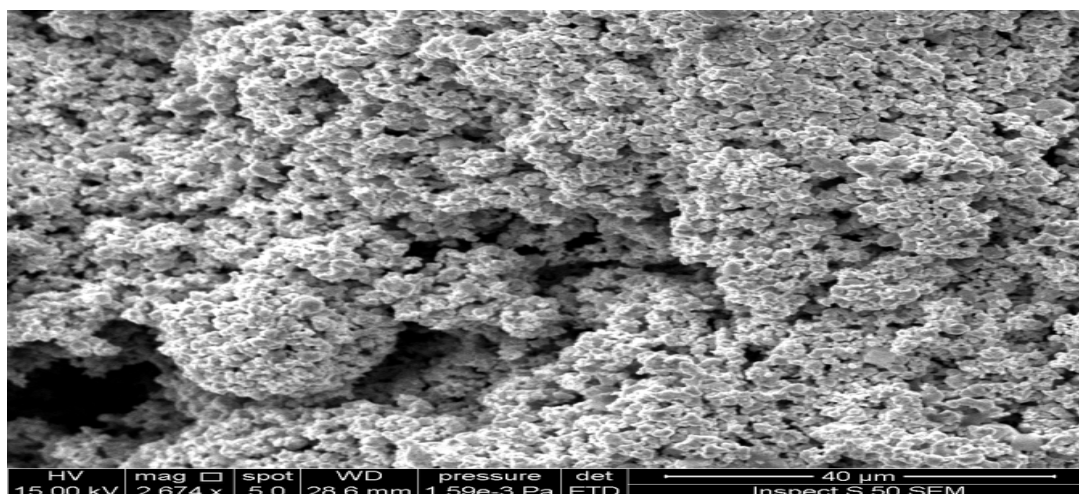


Figure (5): SEM micrographs of silver nanoparticles synthesized by onion extract.

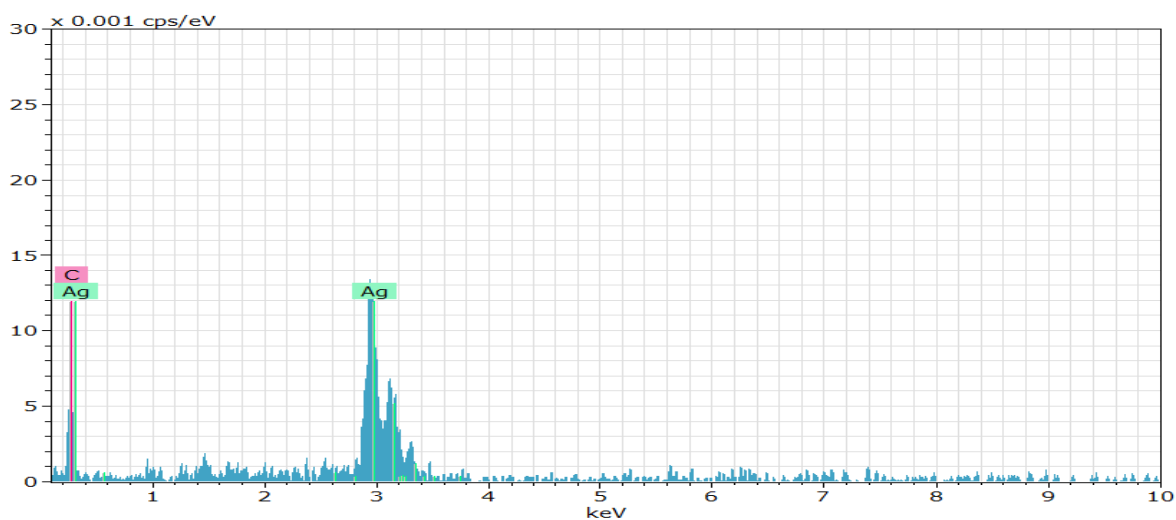


Figure (6): EDX characteristic spectrum obtained for silver powder.

AFM characterization: A  $6.19\mu\text{m} \times 6.19\mu\text{m}$ , AFM image is reported on Figure (7). A statistical treatment of AFM images was performed using specially designed image processing software. To further exploit these measures should be extracted from these images the main geometrical characteristics of these nanostructures. The silver nanoparticles have an average diameter about 28.4nm and an average height 21.9nm. We recognize a very good agreement between the two results: a XRD-size corresponds effectively to a size of AFM.

The data in Table (2) and Figures (8-11) shows that silver nanoparticles synthesised by onion extract has effective antibacterial activities on the test isolates as indicated by the diameter of their zone of inhibition. The inhibition zone was 17mm for *Escherichia coli*, *Enterobacter cloacae*, and *Pseudomonas aeruginosa*, 14mm for *Proteus mirabilis*, 16mm for *Klebsiella oxytoca*, 15mm for *Streptococcus* sp. and *Staphylococcus aureus*, 18mm for *Bacillus* sp. The antimicrobial activity of onion (*Allium cepa*) extract against test bacteria was lower it was between (0-8)mm.

Table (1): Shows the elements in silver nanoparticles

Element	AN	series	[wt.%]	[norm. wt.%]	[norm. at.%]
Carbon	6	K-series	0	0	0
Silver	47	L-series	75.13866	100	100
		Sum:	75.13866	100	100

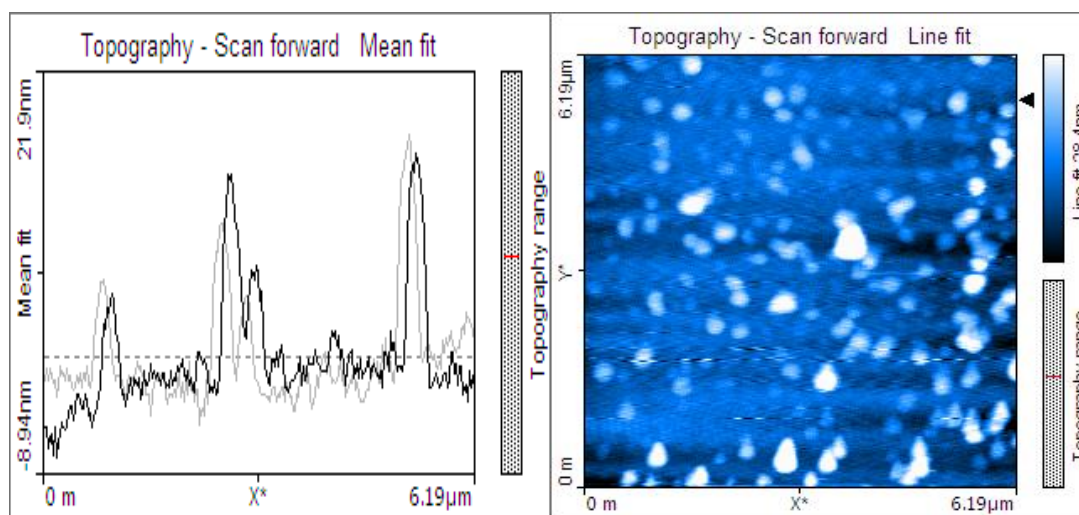


Figure (7): Two-dimensional AFM surface images of Ag NPs colloids deposited on a glass slide.

Table (2): The inhibitory activity of the Ag-NPs synthesis by onion extract against the tested bacteria as demonstrated by diameters of the inhibition zone (mm)\*.

Isolated bacteria	Zone of Inhibition	
	Onion extract	Onion extract/ Ag-NPs
<i>Escherichia coli</i>	7	17
<i>Proteus</i> sp.	6	14
<i>Klebsiella oxytoca</i>	0	16
<i>Streptococcus</i> sp.	6	15
<i>Enterobacter</i> sp.	0	17
<i>Bacillus</i> sp.	0	18
<i>Pseudomonas aeruginosa</i>	0	17
<i>Staphylococcus aureus</i>	8	15

\* Zone of inhibition, including the diameter of the cup plate method (6.0 mm) .The recorded value is mean value of 3 replicates.



Figure (8): The antibacterial effect of Silver nanoparticles synthesis by onion (*Allium cepa*) extract (1), onion (*Allium cepa*) extracts (2) using the test bacterium *Bacillus* sp.



Figure (11): The antibacterial effect of Silver nanoparticles synthesis by onion (*Allium cepa*) extract (1), onion (*Allium cepa*) extracts (2) using the test bacterium *Klebsiella oxytoca*.



Figure (9): The antibacterial effect of Silver nanoparticles synthesis by onion (*Allium cepa*) extract (1), onion (*Allium cepa*) extracts (2) using the test bacterium *Staphylococcus aureus*.



Figure (10): The antibacterial effect of Silver nanoparticles synthesis by onion (*Allium cepa*) extracts (1), onion (*Allium cepa*) extract (2) using the test bacterium *Escherichia coli*.

Silver nanoparticles are nanoparticles of silver which are in the range of 1 and 100nm in size. Silver nanoparticles have unique properties which help in molecular diagnostics, in therapies, as well as in devices that are used in several medical procedures. The major methods used for silver nanoparticle synthesis are the physical and chemical methods. The problem with the chemical and physical methods is that the synthesis is expensive and can also have toxic substances absorbed onto them. To overcome this, the biological method provides a feasible alternative. The major biological systems involved in this are bacteria, fungi, and plant extracts. The major applications of silver nanoparticles in the medical field include diagnostic applications and therapeutic applications.

The synthesis of metal and semiconductor nanoparticles is an expanding research area due to the potential applications for the development of novel technologies. Generally, nanoparticles are prepared by a variety of chemical methods which are not environmentally friendly. We have reported a fast, convenient and extracellular method for the synthesis of silver nanoparticles by reducing silver nitrate with the help of onion (*Allium cepa*) extract. The nanoparticles were characterized using UV-visible, FT-IR, XRD, AFM, and SEM methods. The surface plasmon resonance peaks in absorption spectra for silver colloidal solution showed that the absorption maximum range was at 380-440nm. The functional biomolecules such as carboxyl groups present in the seaweed responsible for the silver nanoparticles formation were characterized by FT-IR. The XRD results suggested that the crystallization of the bio-organic phase occurs on

the surface of the silver nanoparticles or vice versa. The broadening of peaks in the XRD patterns was attributed to particle size effects and the average particles size about 25.17nm. The silver nanoparticles examined by AFM have an average diameter about 28.4nm and an average height 21.9nm. We recognize a very good agreement between the two results: a XRD-size corresponds effectively to a size of AFM.

The results of the present study shows that silver nanoparticles synthesised by onion extract has effective antibacterial activities on the test isolates as indicated by the diameter of their zone of inhibition. The inhibition zone was 17mm for *Escherichia coli*, *Enterobacter cloacae*, and *Pseudomonas aeruginosa*, 14mm for *Proteus mirabilis*, 16mm for *Klebsiella oxytoca*, 15mm for *Streptococcus* sp. and *Staphylococcus aureus*, 18mm for *Bacillus* sp.. Whereas the antimicrobial activity of onion (*Allium cepa*) against test bacteria was lower it was between (0-8)mm Table (2) and Figures (8-11). Our interpretation of these results, the silver nanoparticles synthesized by onion extract has another mechanism to kill bacteria more than onion extract.

The mechanism of the inhibitory effects of Ag ions on microorganisms is partially known. Some studies have reported that the positive charge on the Ag ion is crucial for its antimicrobial activity through the electrostatic attraction between negative charged cell membrane of microorganism and positive charged nanoparticles (Hamouda, *et al.*, 2000; Dibrov, *et al.*, 2002; Dragieva, *et al.*, 1999). In contrast, Sondi and Salopek-Sondi, (2004) reported that the antimicrobial activity of silver nanoparticles on Gram-negative bacteria was dependent on the concentration of Ag nanoparticle, and was closely associated with the formation of 'pits' in the cell wall of bacteria. Then, Ag nanoparticles accumulated in the bacterial membrane caused the permeability, resulting in cell death. However, because those studies included both positively charged Ag ions and negatively charged Ag nanoparticles, it is insufficient to explain the antimicrobial mechanism of positively charged Ag nanoparticles. Therefore, we expect that there is another possible mechanism. Amro *et al.*, suggested that metal depletion may cause the formation of irregularly shaped pits in the outer membrane and change membrane permeability, which is caused by progressive release of lipopolysaccharide molecules and membrane proteins (Amro, *et al.*, 2000). Also, Sondi and Salopek-Sondi speculate that a similar mechanism may cause the degradation of the membrane structure of *E. coli* during treatment with Ag nanoparticles (Sondi and Salopek-Sondi,

2004). Although their inference involved some sort of binding mechanism, still unclear is the mechanism of the interaction between Ag nanoparticles and component(s) of the outer membrane. Recently, Danilczuk and co-workers (2006) reported Ag-generated free radicals through the ESR study of Ag nanoparticles. We suspect that the antimicrobial mechanism of Ag nanoparticles is related to the formation of free radicals and subsequent free radical-induced membrane damage.

Our results support the hypothesis that Ag nanoparticles can be prepared in a simple and cost-effective manner and are suitable for formulation of new types of bactericidal materials.

### Conclusions

This study describes a simple environmentally ecofriendly benign method of synthesis of silver nanoparticles from plants which is the best source. This method can be further used for industrial production of nanoparticles at room temperature and with a single step. This investigation provides evidence that plant extract stabilized nanoparticles may be ideal candidates for future studies exploring their use in biomedical and pharmacy applications. This synthesis procedure offers a less cost-effective and green alternative to traditional protocols that may be readily scaled up for industry as a result of the low synthesis temperatures and time required. Since *A. cepa* (onion) is easily available throughout the nation and also is used in every house for cooking as a flavoring agent, the active nano compound from this can be prepared and used effectively in the field of diagnostic, antimicrobial and therapeutics.

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